

## Monitoring the seasonal cycle of *Emiliana huxleyi* populations in the subantarctic Southern Ocean

### Andrés S. Rigual-Hernández

University of Salamanca, Department of Geology, 37008 Salamanca, Spain; arigual@usal.es

### Tom W. Trull

University of Tasmania, Antarctic Climate and Ecosystems Cooperative Research Centre, Hobart, Tasmania 7001, Australia; CSIRO Oceans and Atmosphere Flagship, Hobart, Tasmania 7001, Australia; Tom.Trull@csiro.au

### José-Abel Flores

University of Salamanca, as above; flores@usal.es

### Scott D. Nodder

National Institute of Water and Atmospheric Research, Wellington 6021, New Zealand; Scott.Nodder@niwa.co.nz

### Ruth Eriksen

CSIRO Oceans and Atmosphere Flagship, as above; University of Tasmania, Institute for Marine and Antarctic Studies, Hobart, Tasmania 7001, Australia; Ruth.Eriksen@csiro.au

### Diana M. Davies

University of Tasmania, as above; CSIRO Oceans and Atmosphere Flagship, as above; Diana.Davies@csiro.au

### Gustaaf M. Hallegraef

University of Tasmania, Institute for Marine and Antarctic Studies, as above; gustaaf.hallegraef@utas.edu.au

### Helen C. Bostock

National Institute of Water and Atmospheric Research, as above; Helen.Bostock@niwa.co.nz

### Francisco J. Sierro

University of Salamanca, as above; sierro@usal.es

### Fatima Abrantes

Portuguese Institute for Sea and Atmosphere, Division of Marine Geology, Lisbon, Portugal; University of Algarve, CCMAR, Marine Science Center, Gambelas Campus, 8005-139 Faro, Portugal; fatima.abrantes@ipma.pt

### Anne-Marie Ballegeer

University of Salamanca, as above; amballegeer@usal.es

### Shramik M. Patil

National Centre for Polar and Ocean Research, Vasco-da-Gama, Goa 403804, India; shramik@ncpor.gov.in

### Aleix Cortina

Institute of Environmental Assessment and Water Research (IDAEA-CSIC), Department of Environmental Chemistry, 08034 Barcelona, Spain; acgqam@idea.csic.es

### Lisa C. Northcote

National Institute of Water and Atmospheric Research, as above; lisapeternorthcote@gmail.com

Anthropogenic CO<sub>2</sub>, which has been accumulating in the oceans since the industrial revolution, decreases carbonate concentrations and the pH of the surface ocean. Polar and subpolar regions can be expected to experience the most severe impacts of ocean acidification. Increased CO<sub>2</sub> concentrations in laboratory cultures often have a negative effect on the physiological processes of the most abundant calcifying phytoplankton species, *Emiliana huxleyi*. However, a few studies have found no trend, or even elevated calcification, in *E. huxleyi*. A possible explanation for this may lie in the genetic diversity in *E. huxleyi*. Laboratory experiments are often focused on single strains, and are unable to reproduce the complexity of natural ecosystem features. Therefore, it is of critical importance to assess the response of *E. huxleyi* populations to changing environmental conditions in their natural habitat.

Here, we report on seasonal variations in the abundance and composition of *E. huxleyi* assemblages that were collected using an autonomous water sampler and four moored sediment traps deployed in the Australian and New Zealand sectors of the subantarctic zone. The combination of morphometric and taxonomic analyses, together with in-situ measurements of environmental parameters, allowed us to monitor, with unprecedented detail, the seasonal cycle of *E. huxleyi* morphotypes in the pelagic waters of the subantarctic zone. Additionally, seasonal changes in coccolith weights were estimated using circularly polarised micrographs. A similar seasonality was found in all the time-series analysed, which suggests that the observed seasonal succession of *E. huxleyi* morphotypes must be a circumpolar feature of the subantarctic zone. Finally, the contribution of other coccolithophore species to carbonate export fluxes is discussed.